



# Hydroformylation of glycols using a rhodium(I)(acac)(CO)<sub>2</sub> catalyst

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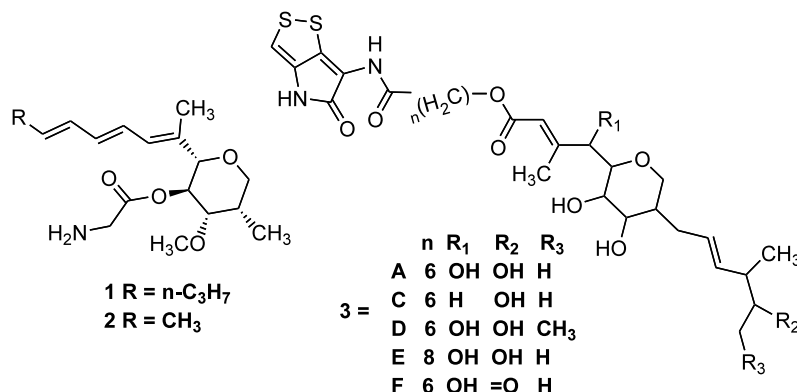
**Abstract**—The hydroformylation of glycols was achieved using Rh(I)(acac)(CO)<sub>2</sub> catalyst. For glucals, the C-2 formyl pyran was the major product whilst for galactal an equimolar mixture of the C-2 and C-1 regioisomers were observed. © 2002 Elsevier Science Ltd. All rights reserved.

C-Branched sugars are essential components in several antibiotics<sup>1</sup> and also key structural subcomponents in many natural products.<sup>2</sup> Members of this group include the polyenes, Restrictin **1** and Lanomycin **2** which have received attention because of their inhibition of P<sub>450</sub> lanosterol C-<sub>14</sub> demethylase and the Thiomarinols D–G **3**, which were recently isolated and have demonstrated potent antimicrobial activity.<sup>3</sup> These classes of compounds are characterized by the presence of a pyran ring which is deoxygenated at C-1 and alkylated at C-2.

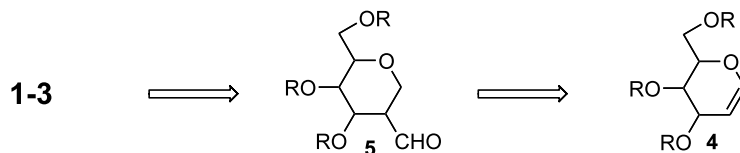
Logical retrosynthetic analysis suggests that a direct route to the pyranose template could involve the regioselective hydroformylation of glycols **4** to their corresponding C-2 intermediates **5**. Intermediate **5** has

the necessary functionality at the C-2 position to allow for conversion to target **1–3** (Scheme 1).

Since its initial description in 1968 by Evans, Osborn and Wilkinson,<sup>4</sup> hydroformylation of alkenes by homogenous rhodium catalysts has attracted the attention of researchers to functionalization of complex molecules.<sup>5</sup> To date, there are only a few reports on hydroformylation of glycols.<sup>6,7</sup> We recently synthesized carba-D-fructofuranose via a hydroformylation of a functionalized cyclopentene.<sup>8</sup> Utilizing the catalyst Rh(acac)(CO)<sub>2</sub> under similar reaction conditions, glycols gave a mixture of the C-1 formyl 2-deoxy C-glycoside and the corresponding C-2 formyl pyran.<sup>9</sup> Thus hydroformylation of tri-*O*-benzyl-D-glucal **6** gave the



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Scheme 1.

formyl adducts **7:8** in ratio of 8:92.<sup>7</sup> Use of the corresponding acetyl protected glucal **9** gave similar results. The 3,4-di-*O*-acetyl-6-deoxy glucal **12** gave only the C-2 formyl product **14**. However, tri-*O*-benzyl-*D*-galactal **15** was quantitatively hydroformylated to give a 1:1 mixture of formyl compounds **16** and **17** (Scheme 2).

The regioselectivity of formyl addition depends on the following factors; polarization of the olefin, relative stability of the alkyl-metal complexes, the difficulty of producing the  $\beta$ -elimination process in conformationally rigid substrates, and the ratio between the rates of formation of the acyl-metal intermediates.<sup>7b</sup> Our results indicate that there is a general tendency to hydroformylate at the C-2 position. Of note, galactal where C3, C4 and C5 positions are *syn* gave 45% of the C-1 formyl regioisomer compared to its glucal counterpart (0–14%). It seems that this difference in stereochemistry at the C-4 position influences some of the above ‘factors’ to favor the C-1 formylation. Thus for galactal, the hydroformylation represents a direct entry into the 2-deoxy-C-glycosides **5**. For all the C-2 regioisomers obtained, we observed minimal elimination suggesting an anti addition to the C-3 substituent. Also no hydrogenation products were observed. The reaction proceeds under neutral conditions and does not need the addition of auxiliary ligands such as  $\text{PPh}_3$  or  $\text{P}(O\text{-}o\text{-}$

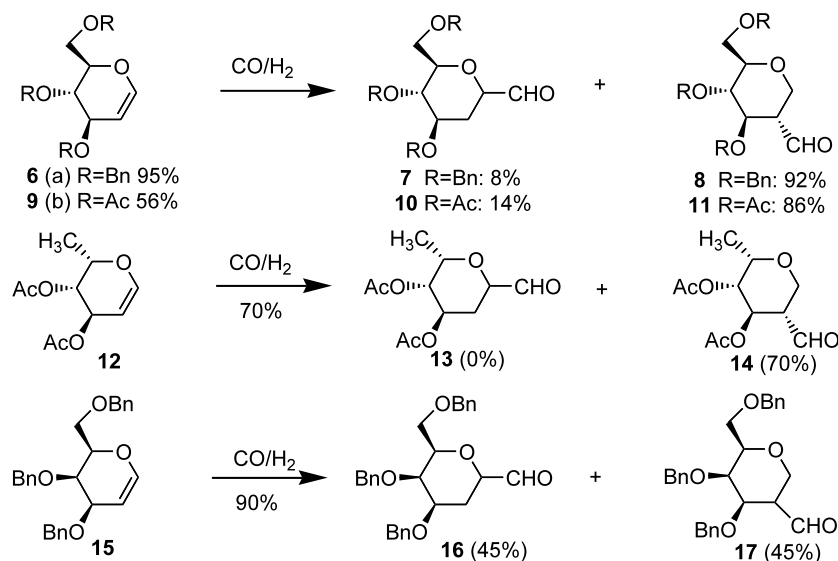
$\text{BuC}_6\text{H}_4)_3$ ,<sup>10</sup> Of note, the use of the  $\text{Co}_2(\text{CO})_8$  under the same reaction conditions gave no reaction.<sup>6</sup>

The regiochemistry of our products was established by comparison to the available literature.<sup>7b</sup> In general, key chemical shift differences exist between the C-1 and C-2 formyl diastereomers. In  $^1\text{H}$  NMR spectra the presence of a ABq, at 3.4 ppm and the absence of any signals upfield to this are indicative of the formyl group at C-2. In  $^{13}\text{C}$  NMR, the presence of an additional signal in the 50–60 ppm region and the absence of a signal upfield to this indicates C-2 formyl attachment.<sup>9</sup>

At present, an extension of this methodology using external chiral ligands is underway in an attempt to direct the regiochemistry to C-1 formyl glycosides.<sup>10</sup> Overall, using the above catalyst, we have demonstrated an efficient and versatile one carbon extension of glycals which allows access to a variety of novel synthetic intermediates for antifungal and antibiotic agents.

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Scheme 2.

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- A solution containing the dicarbonylacetylacetonato rhodium (0.035 g, 0.14 mmol), toluene (70 mL) and the olefin **15** (3.0 g, 1.2 mmol) were placed in a 300 mL mechanically-stirred steel autoclave (equipped with a gas entrainment impeller) which was then charged with carbon monoxide (50 bar) and hydrogen (50 bar). The reaction mixture was stirred at 100°C for 48 h. At the end of this period, the remaining gases were vented off and the solvent was evaporated to furnish the crude aldehydes **16/17** in 1:1 ratio and 90% yield with  $R_f$ s 0.30 and 0.4 (25% EtOAc:PE), respectively. Selected spectroscopic data are as follows; Compound **16**:  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 270 MHz)  $\delta$  2.25 (m, 2H), 3.45–3.61 (m, 3H), 3.80 (m, 3H), 4.35 (t,  $J=7.5$  Hz, 1H), 4.42–4.62 (m, 6H), 4.89 (m, 1H), 7.3 (m, 15H), 9.75 (s, 1H).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 67.5 MHz)  $\delta$  25.4, 68.9, 70.8, 73.2, 73.6, 74.0, 74.9, 75.6, 127.5, 127.8–128.7 (several signals), 138.0, 138.2, 138.5, 203.8. MS (ES)  $m/z$  ( $M+K^+$ ), 485 (base peak), 181. Compound **17**:  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 270 MHz)  $\delta$  3.46 (ABq,  $J=8.9$  Hz,  $\Delta\delta=0.10$  ppm, 2H), 3.59 (m, 3H), 3.80 (dd,  $J=2.2, 8.4$  Hz, 1H), 4.00 (d,  $J=2.2$  Hz, 1H), 4.13 (d,  $J=7.0$  Hz, 1H), 4.47 (ABq,  $J=11.9$  Hz,  $\Delta\delta=0.08$  ppm, 2H), 4.60 (ABq,  $J=11.4$  Hz,  $\Delta\delta=0.22$  ppm, 2H), 4.76 (ABq,  $J=11.6$  Hz,  $\Delta\delta=0.31$  ppm, 2H), 7.3 (m, 15H), 9.84 (s, 1H).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 67.5 MHz)  $\delta$  50.1, 65.6, 69.2, 71.3, 71.5, 73.7, 74.7, 77.8, 79.2, 127.9–128.7 (several signals), 137.4, 137.8, 138.4, 202.8. MS (ES)  $m/z$  ( $M+Na^+$ ), 469 (base peak), 181.
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